

UNIVERSIDADE FEDERAL DE PELOTAS

Programa de Pós-Graduação em Biotecnologia



Dissertação

**CLONAGEM DO GENE QUE CODIFICA PARA A LISTERIOLISINA EM
BCG $\Delta leuD$ E AVALIAÇÃO DA ATIVIDADE ANTITUMORAL EM
CÉLULAS DE CÂNCER DE BEXIGA**

KAREN SILVA LEAL

Pelotas, 2013

KAREN SILVA LEAL

**CLONAGEM DO GENE QUE CODIFICA PARA A LISTERIOLISINA EM BCG
ΔleuD E AVALIAÇÃO DA ATIVIDADE ANTITUMORAL EM CÉLULAS DE
CÂNCER DE BEXIGA**

Dissertação apresentada ao
Programa de Pós-Graduação em
Biotecnologia da Universidade
Federal de Pelotas, como requisito
parcial à obtenção do título de
Mestre em Ciências (área do
conhecimento: Biotecnologia)

Orientador: Prof Dr. Odir Antônio Dellagostin

Coorientadora: Prof Dr. Sibeles Borsuk

Pelotas, 2013

Dados Internacionais de Publicação (CIP)

L433c Leal, Karen Silva

Clonagem do gene que codifica para para a listeriolisina em BCG e avaliação da atividade antitumoral em células de câncer de bexiga / Karen Silva Leal; Odir Antônio Dellagostin, orientador; Sibeles Borsuk, co-orientador. - Pelotas, 2013. 42 f.

Dissertação (Mestrado em Biotecnologia), Programa de Pós-Graduação em Biotecnologia, Universidade Federal de Pelotas. Pelotas, 2013.

1. Mycobacterium Bovis . 2. Biotecnologia. 3. Listeriolisina. 4. câncer de bexiga. I. Dellagostin, Odir Antônio. II. Borsuk, Sibeles. III. Título.

CDD: 620.8

Catálogo na Fonte: Leda Cristina Peres Lopes CRB:10/2064
Universidade Federal de Pelotas

Banca examinadora:

Prof.^a Dr. Fabiana Kömmling Seixas, Universidade Federal de Pelotas

Dr^a Daiane Hartwig, Universidade Federal de Pelotas

Dr^a Daniela Ramos, Universidade Federal do Rio Grande

Prof. Dr. Odir Antônio Dellagostin, Universidade Federal de Pelotas

Agradecimentos

À Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) pela concessão da bolsa de mestrado.

Ao Programa de Pós-Graduação em Biotecnologia e à Universidade Federal de Pelotas pelos conhecimentos adquiridos.

Ao meu orientador, Odir Antônio Dellagostin, pela orientação, oportunidade e confiança no meu trabalho.

A minha coorientadora, Sibele Borsuk, pela incansável ajuda, orientação e disponibilidade.

Aos meus pais, Nilva e Valdeci, pelo apoio, incentivo e lutas para concretização dos meus sonhos.

Ao meu irmão Willian Leal, uma das pessoas mais especiais na minha vida.

A Bianca Leal, minha irmã de mãe diferente, amiga, companheira.

Ao meu amigo de todas as horas, Sérgio Jorge, uma pessoa maravilhosa, única, só tenho a agradecer por ele fazer parte da minha vida.

A Caroline Rizzi, pelo companheirismo e por todo o conhecimento transmitido.

A Karine Begnini, pela disponibilidade e paciência na execução dos nossos experimentos.

A Ana Carolina Peiter pela lealdade, dedicação, competência e responsabilidade.

A Andrea Rezende, pelo carinho, amizade, sinceridade, e conselhos.

Aos meus amigos de longa data, Vagner Madruga, Débora Bisognin, Thays Garcia, Kathleen Winkel, Taiane Camacho e Lidiane Bevilacqua “A amizade desenvolve a felicidade e reduz o sofrimento, duplicando a nossa alegria e dividindo a nossa dor”. Isso que sinto por cada um de vocês.

A Flávia Vargas, por todas as orações e por estar sempre na torcida pelo meu sucesso.

A todos os colegas e amigos do Laboratório de Vacinologia, pelo convívio todo dia, pelos aprendizados, e por tornarem o ambiente de trabalho mais agradável.

A todos que não foram mencionados aqui, mas que de alguma forma foram importantes e contribuíram para mais uma etapa na minha vida.

A Deus por guiar meu caminho e minha intuição.

Muito Obrigada!

RESUMO

LEAL, Karen. **CLONAGEM DO GENE QUE CODIFICA PARA A LISTERIOLISINA EM BCG $\Delta leuD$ E AVALIAÇÃO DA ATIVIDADE ANTITUMORAL EM CÉLULAS DE CÂNCER DE BEXIGA** 2013. 42f. Dissertação (Mestrado) - Programa de Pós-Graduação em Biotecnologia. Universidade Federal de Pelotas, Pelotas.

O *Mycobacterium bovis* BCG é utilizado mundialmente como vacina contra tuberculose há mais de meio século, com altos níveis de segurança. O BCG possui características que o tornam um promissor vetor para a produção de vacinas recombinantes multivalentes. Modificações genéticas podem aprimorar as propriedades imunogênicas e adjuvantes do BCG. Uma dessas modificações permite que o BCG escape do endossoma de células apresentadoras de antígenos. Desta maneira, os imunógenos de BCG acessam vias de apresentação cruzada de antígenos, ampliando a sua imunogenicidade. Este trabalho teve como objetivo desenvolver uma cepa de BCG $\Delta leuD$ expressando listeriolisina (Hly) de *Listeria monocytogenes* e avaliar seu potencial citotóxico na linhagem celular 5637 de câncer de bexiga. Para isso, o BCG $\Delta leuD$ foi transformado com o plasmídeo recombinante pUP410 contendo o gene *hly*. A cepa obtida demonstrou um nível de citotoxicidade similar à cepa BCG Pasteur parental, quando avaliada na linhagem celular de câncer de bexiga. Esta cepa necessita ser avaliada em outras linhagens de câncer para comprovar seu possível efeito citotóxico potencializado.

Palavras chaves: *Mycobacterium bovis* BCG , $\Delta leuD$, listeriolisina, câncer de bexiga.

ABSTRACT

LEAL, Karen. **CLONING OF THE GENE THAT CODES FOR LISTERIOLYSIN ON BCG Δ leuD AND EVALUATION OF THE ANTITUMORAL ACTIVITY ON BLADDER CANCER CELLS** 2013. 42f. Dissertação (Mestrado) - Programa de Pós-Graduação em Biotecnologia. Universidade Federal de Pelotas, Pelotas.

Mycobacterium bovis BCG has been used worldwide as a vaccine against tuberculosis for over fifty years with an outstanding safety record. BCG has features that make it a promising vector for production of recombinant multivalent vaccines. Genetic modification can enhance the adjuvant and immunogenic properties of BCG. One of these modifications would allow BCG to escape the endosome of antigen presenting cells. Therefore, the immunogens of BCG can access antigens cross priming routes, increasing their immunogenicity. The aim of this work was to develop a strain of BCG Δ leuD expressing listeriolysin (Hly) of *Listeria monocytogenes* and evaluate its cytotoxic potential on cell line 5637 of bladder cancer. Thus, BCG Δ leuD was transformed with recombinant plasmid pUP410 containing the *hly* coding sequence. The recombinant BCG strain expressing listeriolysin showed a similar level of cytotoxicity when compared to the parental BCG Pasteur. This strain needs to be evaluated in other cancer lines in order to confirm its possible potentiated cytotoxic effect.

Keywords: *Mycobacterium bovis* BCG, Δ leuD, listeriolysin, bladder cancer;

LISTA DE FIGURAS

- Figura 1. Mecanismo do BCG recombinante expressando listeriolisina. 15
- Figura 1. Representação esquemática do gene sintético clonado no 32
vetor pUP410.
- Figura 2. Eletroforese em gel de agarose confirmando a detecção do 33
mRNA.
- Figura 3. BCG $\Delta leuD/hly$ inibindo o crescimento de células de câncer de 34
bexiga após 48 h de tratamento.
- Figura 4. Citometria de fluxo demonstrando o número de células em 35
apoptose inicial e tardia.

LISTA DE ABREVIATURAS E SIGLAS

BAAR- Bacilo Alcool-Ácido Resistente

BCG - Bacillus Calmette-Guérin

rBCG – BCG recombinante

Hly- Listeriolisina

HPV- Human Papiloma Virus

PSA - Prostate Specific Antigen

RT – PCR - Reverse-Transcription Polymerase Chain Reaction

S1PT- Subunidade 1 da toxina de Bordetella pertussis

TB- Tuberculose

SUMÁRIO

1. Introdução.....	10
1.1 <i>Mycobacterium bovis</i> BCG.....	10
1.2 BCG recombinante.....	11
1.3 Sistema auxotrófico $\Delta leuD$	12
1.4 Imunoterapias e câncer.....	13
1.5 Listeriolisina e câncer.....	15
2. Objetivos.....	17
2.1 Geral.....	17
2.2 Específicos.....	17
3. Artigo.....	18
3.1 Abstract.....	20
3.2 Introduction.....	21
3.3 Material and Methods.....	22
3.4 Results.....	25
3.5 Discussion.....	27
3.6 References.....	30
4. Conclusões.....	36
5. Referências.....	37

1. INTRODUÇÃO

O *Mycobacterium bovis* bacilo Calmette-Guérin (BCG) é um bacilo álcool-ácido resistente (BAAR) não esporulado, do grande grupo das micobactérias (PARISH; STOKER, 1998). Sua parede celular é composta majoritariamente por lipídeos, o que lhe proporciona muitas das propriedades antigênicas, adjuvantes, anti-tumorais e de virulência (FORRELLAD et al., 2013). Apesar disso, as micobactérias não possuem fatores de virulência clássicos, como toxinas, que são típicas de outros patógenos bacterianos. Dessa maneira, outros fatores de virulência como enzimas de várias rotas lipídicas, proteínas de superfície celular, sistemas de transdução de sinal e proteínas reguladoras permitem sua sobrevivência no interior dos macrófagos, como a inibição da fusão do fagossomo com o lisossomo e escape do ambiente ácido (FORRELLAD et al., 2013).

A utilização do BCG como vacina foi iniciada em 1921 e, desde então, mais de três bilhões de pessoas já foram vacinadas, sendo considerada atualmente a vacina mais administrada no mundo (BLOOM; FINE, 1994). Em 2011 a Organização Mundial da Saúde estimou 8,7 milhões de casos de tuberculose no mundo (WHO; 2012). O BCG é a única vacina usada na prevenção da tuberculose (TB) em humanos (VON REYN; VUOLA, 2002) e oferece vantagens únicas: não é afetado por anticorpos maternos; pode ser administrada logo após o nascimento; uma dose única é segura, estável, possui baixo custo na produção em relação a outras vacinas vivas e confere uma imunidade prolongada (STOVER et al., 1991; (BASTOS et al., 2009a).

1.2 BCG recombinante

Avanços na área da tecnologia do DNA recombinante, bem como o conhecimento do genoma e o desenvolvimento de sistemas de expressão de antígenos heterólogos em BCG, fizeram com que essa bactéria se tornasse um excelente candidato a vetor de vacina recombinante multivalente expressando antígenos heterólogos de outros microrganismos (JACOBS et al., 1987; (BASTOS et al., 2009b). A vacina com rBCG confere vantagens em relação ao BCG selvagem, como uma maior segurança, além de causar menos efeitos colaterais e ser capaz de se manter a longo prazo no organismo inoculado (WANG et al., 2012). A indução de uma resposta imune contra antígenos heterólogos pela inoculação de rBCG e a primeira evidência de proteção foram descritas por STOVER et al.,1991.

Desde então, inúmeros outros estudos confirmam o uso do rBCG contra agentes parasitários, bactérias, e antígenos virais (BASTOS et al., 2009b). Para a obtenção de uma resposta protetora satisfatória, a via de administração é um fator importante para garantir o sucesso na vacinação, pois a imunogenicidade de algumas vacinas é alterada quando não for administrada adequadamente (VARALDO et al., 2004).

A construção de cepas recombinantes que proporcionem maior estímulo do sistema imune e aumento do efeito antitumoral da bactéria constitui um dos principais focos de pesquisas em melhorias imunoterapêuticas do BCG (AMIRKHAH et al., 2009). A utilização dos rBCG para antígenos heterólogos de diferentes bactérias, bem como a expressão de combinados antigênicos, também demonstram eficácia aumentada frente a tumores, tanto em ensaios *in vitro* quanto em modelos animais (ANDRADE et al., 2010;CHUNG et al.,

2003).O rBCG tem sido, sem dúvida, uma das terapias com mais sucesso diante de tumores de bexiga (HERR et al., 2008).

1.3 Sistema auxotrófico $\Delta leuD$

Um mutante de BCG Pasteur denominado BCG $\Delta leuD$ foi desenvolvido por nosso grupo de pesquisa. Esse sistema de seleção é baseado no uso de uma cepa auxotrófica para o aminoácido leucina e na complementação com o gene *leuD* inserido no vetor plasmidial pUP410. Esse sistema possui vantagens como: promover uma seleção *in vivo* ativa, não necessitando de vetores que contenham genes de resistência a antibióticos (BORSUK et al., 2007).

Estudos realizados por (BANGE; BROWN; JACOBS, Jr., 1996) mostraram que o crescimento de *M. bovis* BCG auxotrófico para leucina era restrito no interior de macrófagos. Quando este mutante foi complementado com o gene *leuD* de *Escherichia coli* os níveis de crescimento no interior dos macrófagos foram restabelecidos, demonstrando a eficiência desse sistema de seleção. Um estudo prévio comprovou também a eficácia de um BCG $\Delta leuD$ expressando o antígeno 85B quando submetido a um teste desafio com bovinos. Este se manteve de forma estável mesmo sem a presença de um marcador de resistência a antibiótico (RIZZI et al., 2012). Segundo (BEGNINI et al., 2013) este BCG $\Delta leuD/Ag85B$ também se mostrou eficaz na imunoterapia atuando na inibição do crescimento de células de câncer de bexiga.

1.4 Imunoterapias e câncer

A imunoterapia tem como objetivo a estimulação do sistema imune, alterando a resposta biológica mediada por células ou interferindo na interação antígeno-anticorpo, sendo esta uma promissora estratégia potencialmente eficaz para o tratamento do câncer (ALDRICH et al., 2010). O sistema imunológico é capaz de reconhecer e extinguir lesões pré-cancerosas e cancerosas (SHENG; HUANG, 2011). Esse reconhecimento é obtido principalmente através de vacinação com peptídeos antigênicos ou através de células dendríticas ativadas (DRANOFF, 2004).

Desse modo, o uso do BCG para terapia de câncer foi proposto em 1930. No entanto pouco foi feito até os anos 1950 e 1960, quando foram diagnosticados outros tipos de tumores, como leucemia, câncer colo-retal, câncer de pulmão e melanomas. Os resultados não impressionavam muito, exceto por um estudo promissor de Mathé no tratamento de leucemia linfoblástica em 1969 que outros pesquisadores não tinham confirmado até então (LOCKYER et al., 2001). O advento da quimioterapia e radioterapia reduziu o entusiasmo com o BCG. Enquanto isso, em 1966, estudos de Coe & Feldman mostraram uma reação de hipersensibilidade na bexiga de cobaias ao serem administrados com BCG. Com isso, foram capazes de explorar a resposta imune quando obtiveram o êxito no tratamento câncer (LOCKYER et al., 2001).

A utilização do BCG na imunoterapia para câncer de bexiga teve início baseando-se principalmente em três razões: no tratamento de *carcinoma-in-situ*; na redução da frequência de tumores recorrentes e também na profilaxia da doença (KAWAI et al., 2013). A partir de então, muitas pesquisas tem se

mostrado eficientes nas imunoterapias contra vários tipos de câncer (WALLECHA et al., 2011; KAWAI et al., 2013).

NASCIMENTO e colaboradores, construíram uma cepa de BCG recombinante em fusão com subunidade 1 da toxina de Bordetella pertussis (S1PT) e demonstraram que essa vacina é capaz de estimular uma resposta imune Th1 mais forte que o BCG selvagem. Em outro estudo *in vivo*; (ANDRADE et al., 2010) confirmam que no ensaio de citotoxicidade, camundongos tratados com rBCG-S1PT foram mais eficazes em destruir células da linhagem MB49 de tumores de bexiga. É provável que as citocinas secretadas tenham ativado uma maior produção de linfócitos e células efectoras durante o contato do rBCG com células tumorais. Esses estudos também constatarem uma redução significativa em tumores, aumentando assim a taxa de sobrevivência dos camundongos em relação ao grupo controle vacinados somente com BCG.

Em outro trabalho, a utilização de uma cepa recombinante de BCG superexpressando o antígeno 85B também se mostrou eficiente em ensaios citotóxicos com a linhagem de carcinoma de bexiga 5637. Foi constatado um aumento da morte celular quando estas foram tratadas com BCG Δ euD/ag85B em relação a células tratadas apenas com o controle BCG Pasteur que é usado no tratamento convencional (BEGNINI et al., 2013). Estes dados comprovam a eficiência gerada pelo rBCG, enfatizando seu uso na imunoterapia de câncer de bexiga, aumentando as perspectivas do emprego de cepas geneticamente modificadas para aprimorar o tratamento já existente.

1.5 Listeriolisina e câncer

A bactéria *Listeria monocytogenes* vem se destacando na imunoterapia para tumores, como o câncer de próstata, câncer de colo de útero, fazendo uso de modificações genéticas para a expressão de antígenos heterólogos (SHAHABI et al., 2008 WALLECHA et al., 2011). A *L. monocytogenes* é atrativa para o desenvolvimento de imunoterapias, pois gera uma forte resposta imune inata e adaptativa. Ela é uma bactéria intracelular que acomete macrófagos e células dendríticas. Quando secreta uma proteína nativa, a listeriolisina consegue formar poros no fagolisossomo, escapando para o citosol, onde consegue acessar outras vias de apresentação de antígenos, ampliando sua imunogenicidade (WALLECHA et al., 2011). Alterações genéticas no BCG expressando listeriolisina podem ser alternativas para aprimorar as imunoterapias de câncer já existentes. Na Figura 1 está representado o mecanismo de ação do BCG recombinante expressando a listeriolisina:

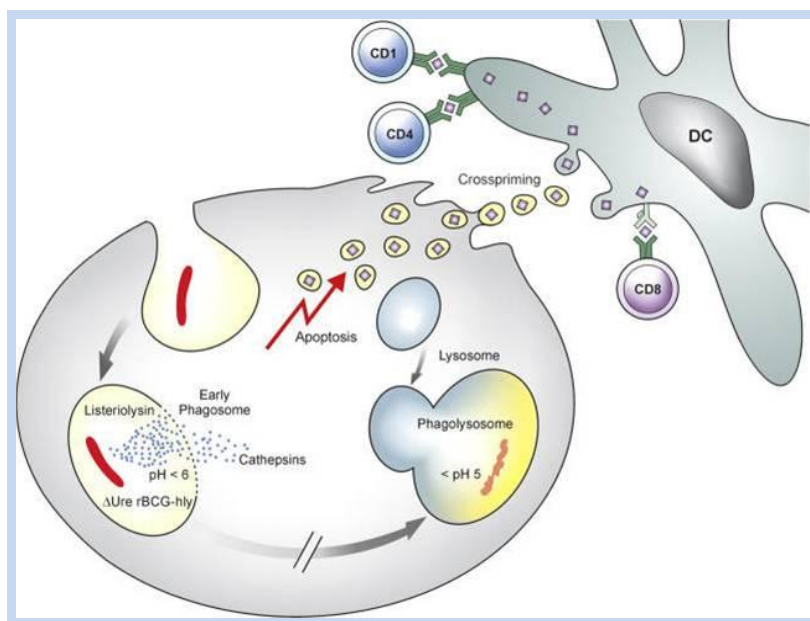


Figura 1: Descrição esquemática do sistema empregando BCG recombinante expressando listeriolisina, acessando outras vias de apresentação de antígenos, aumentando a imunogenicidade da vacina (Imagem concedida por Stefan Kaufmann).

Outras propriedades da *L. monocytogenes* indicam que quando fusionados com uma forma truncada da listeriolisina O, antígenos expressados e secretados pela *L. monocytogenes* foram significativamente mais imunogênicos. Em estudos prévios, uma cepa recombinante de *L. monocytogenes* foi capaz de causar a regressão de determinados tumores como em câncer de próstata (SHAHABI et al., 2008). Também foi avaliado o potencial de listeriolisina O para erradicar tumores induzidos por HPV (WALLECHA et al., 2011).

2. OBJETIVO

2.1 Geral

Construir uma cepa de *M. bovis* BCG Δ leuD recombinante contendo o gene que codifica para a listeriolisina de *L. monocytogenes* e avaliar seu potencial citotóxico na linhagem 5637 de células de câncer de bexiga.

2.2 Específicos

- 1- Obter a sequência codificadora da listeriolisina e clonar no vetor de expressão pUP410;
- 2- Construir a cepa recombinante e confirmar a expressão da listeriolisina por RT-PCR;
- 3- Avaliar o potencial da cepa recombinante em ensaios de citotoxicidade em linhagens de células de câncer de bexiga

3. ARTIGO

**CLONING OF THE GENE THAT CODIFIES FOR LISTERIOLYSIN ON BCG
*Δ*leu*D* AND EVALUATION OF THE ANTITUMORAL ACTIVITY ON
BLADDER CANCER CELLS**

(Artigo científico escrito sob formato do periódico Current Microbiology)

CLONING OF THE GENE THAT CODIFIES FOR LISTERIOLYSIN ON BCG *ΔleuD* AND EVALUATION OF THE ANTITUMORAL ACTIVITY ON BLADDER CANCER CELLS

¹Karen Silva Leal, ¹Caroline Rizzi, ²Sibele Borsuk, ¹Sérgio Jorge,³Karine Begnini,

³Fabiana Seixas ³Tiago Collares ¹Ana Carolina Peiter and ¹ Odir Antonio Dellagostin

¹Laboratório de Vacinologia, Núcleo de Biotecnologia, Centro de Desenvolvimento Tecnológico (CDTec): Karen Silva Leal, karensleal@hotmail.com; Caroline Rizzi, ccrizzi@yahoo.com.br; Sérgio Jorge, sergiojorgevet@hotmail.com; Ana Carolina Peiter, acpeiter@gmail.com; Odir Antonio Dellagostin, odir@terra.com.br;² Laboratório

de pesquisa em doenças infecciosas, Núcleo de Biotecnologia, Centro de Desenvolvimento Tecnológico (CDTec): Sibele Borsuk sibeleborsuk@gmail.com;

³Laboratório de Genômica Funcional, Karine Rech Begnini, karibegnini@gmail.com; Fabiana Kömmling Seixas, seixas.fk@gmail.com, Tiago Collares, collares.t@gmail.com

Universidade Federal de Pelotas, Campus Universitário Capão do Leão, CP 354, CEP 96010-900, Pelotas city, RS, Brazil.

¹Corresponding author: Odir Antonio Dellagostin, Laboratório de Vacinologia, Núcleo de Biotecnologia, Centro de Desenvolvimento Tecnológico, Universidade Federal de Pelotas, Campus Capão do Leão, 96010-900, Pelotas city, RS, Brazil. Tel: +55 53 32757583. Fax: +55 53 32757551. E-mail address: karensleal@hotmail.com

ABSTRACT

Mycobacterium bovis BCG has been used worldwide as a vaccine against tuberculosis for over fifty years with an outstanding safety record. BCG has features that make it a promising vector for production of recombinant multivalent vaccines. Genetic modification can enhance the adjuvant and immunogenic properties of BCG. One of these modifications would allow BCG to escape the endosome of antigen presenting cells. Therefore, the immunogens of BCG can access antigens cross priming routes, increasing their immunogenicity. The aim of this work was to develop a strain of BCG Δ leuD expressing listeriolysin (Hly) of *Listeria monocytogenes* and evaluate its cytotoxic potential on cell line 5637 of bladder cancer. Thus, BCG Δ leuD was transformed with recombinant plasmid pUP410 containing the *hly* coding sequence. The recombinant BCG strain expressing listeriolysin showed a similar level of cytotoxicity when compared to the parental BCG Pasteur. This strain needs to be evaluated in other cancer lines in order to confirm its possible potentiated cytotoxic effect.

Keywords: *Mycobacterium bovis*, Δ leuD, listeriolysin, bladder cancer;

1. INTRODUCTION

The *Mycobacterium bovis* bacillus Calmette-Guérin (BCG) strain is used worldwide as a vaccine against tuberculosis [3]. It has also been used as a vector for the development of recombinant and multivalent vaccines against many infectious diseases and cancer immunotherapy [14,16,4]. BCG has adjuvant properties, is not affected by maternal antibodies, a single dose confers a long immune response, is stable and safe, can be administered orally and has a low production cost compared to other live vaccines [14].

Various expression systems for exogenous antigens in BCG have been developed to enable the construction of recombinant BCG (rBCG) [19]. Most of these systems employs integrating vectors or episomal vectors containing antibiotics resistance genes as selection markers, which respectively confer low levels of protein expression and absence of in vivo selection, hence low vector stability [6,3]. Our research group has developed a BCG $\Delta leuD$ expression system, which allows for stable genetic transformation of exogenous genes without the use of antibiotics as selection markers. This system employs the complementary auxotrophic selection leucine marker, allowing the production of safe and stable recombinant strains, in vivo, resulting in high levels of protein expression [5].

An improvement on the ability of the current BCG to induce a broader immune response can be achieved by allowing the escape of the BCG from the endosome of antigen-presenting cells. This results in cross-presentation of antigens, increasing their immunogenicity [13,16]. The listeriolysin (Hly) is secreted by the intracellular bacterium *Listeria monocytogenes* and is responsible for the release of this

microorganism from the phagosomes into the cytoplasm of infected cells [8]. Besides its use as a vaccine, BCG has also been exploited in anti-tumour therapy. BCG expressing foreign antigens or overexpressing some of its own antigens is a promising approach to improve the performance of BCG anti-tumour therapy [2]. Current studies are developing rBCG strains to further improve the effectiveness of the therapy [4,7].

The aim of this study was to construct a BCG Δ *leuD*/hly secreting listeriolysin and to evaluate the cytotoxic effect in 5637 bladder carcinoma cells line.

2. MATERIALS AND METHODS

2.1 Coding sequence of the listeriolysin and primer design

A synthetic gene containing the coding sequence for the Hly protein was constructed based on the sequence deposited in GenBank (NC 003210) derived from *Listeria monocytogenes* EGD-e. To facilitate secretion of the protein by BCG, the synthetic gene had a region corresponding to the listeriolysin signal peptide replaced by Ag85B signal peptide, a protein normally secreted by *M. bovis*. The primers used in PCR amplification were (5'-ACTGGGGTACCGGCCGGTGAAGTTC-3' and 5'-GGGGTACCGGGTGC GCGGTCTGCCA-3'), designed using *Vector NTI* 11.0 (Invitrogen TM, Carlsbad, CA, EUA).

2.2 Cloning of the hly gene into the expression vector

The PCR product was digested with *KpnI* enzyme (Promega), and cloned into pUP410 mycobacterial expression vector [5], which had previously been digested with the same restriction enzyme (Figure 1). Competent *E. coli* was transformed with

ligation product and a recombinant plasmid (pUP410/hly) was identified and verified by restriction enzyme digestion and PCR.

2.3 Construction of rBCG Δ leuD containing *hly*

Electrocompetent BCG Pasteur Δ leuD was transformed with pUP410/hly and grown on Middlebrook 7H9 medium (Difco Laboratories, Detroit, MI, USA) supplemented with 0.5% glycerol, 0.05% Tween 80, 10% oleic albumin-dextrose-catalase (OADC) and supplemented with 100 mg/ml L-leucine (Sigma). The recombinant BCG was grown to an optical density at 600 nm (OD₆₀₀) of 0.6 in selective 7H9 media, the culture (10 mL) was centrifuged (4,000g for 10 min). The detection of mRNA was confirmed by Reverse-Transcription PCR (RT-PCR). BCG Δ leuD/hly was grown in liquid culture and then tested regarding its cytotoxicity on a bladder carcinoma cell line.

2.4 Reverse-Transcription PCR (RT-PCR)

The *hly* expression profiles was investigated by RT-PCR. Total RNA was isolated with TRIzol® Reagent (Invitrogen™, Carlsbad, USA). DNase treatment of RNA samples was conducted with a DNA-free® Kit (Ambion™, USA) following the manufacturer's protocol. First-strand cDNA was performed with 2µg of RNA using SuperScript™ III Reverse Transcriptase (Invitrogen™, Carlsbad, USA) according to the manufacturer's protocol. RT-PCR reaction was run using PCR Master Mix (Applied Biosystems, UK) and the primers used were (5'- GCAAGCCTACCCGAATGTGT- 3' and 5'GTCGGTTCGTTGACGTTGAC-3').

2.5 Cell culture

The human bladder carcinoma cell line 5637 was obtained from the Rio de Janeiro Cell Bank (PABCAM, Federal University of Rio de Janeiro, RJ, Brazil). Cells were grown at 37 °C in an atmosphere of 95% humidified air and 5% CO₂. The cells were cultured in Dulbecco's modified Eagle's medium (DMEM), supplemented with 10% fetal bovine serum (FBS). The experiments were performed with cells and bacteria in the logarithmic phase of growth and all experiments were performed in triplicate.

2.6 Determination of cytotoxicity

The viability of the 5637 cell line was determined by measuring the reduction of soluble MTT [3-(4,5-dimethylthiazol- 2-yl)-2,5-diphenyltetrazolium bromide] to water insoluble formazan [1]. Cells were seeded at a density of 2×10^4 cell per well in a volume of 100 μ L in 96-well plates, and grown at 37 °C in a 5% CO₂ atmosphere for 24 h before being used in the cell viability assay. Cells were then incubated with the BCG strain (BCG Δ leuD- BCG Δ leuD/hly1, BCG Δ leuD/hly2 and BCG Δ leuD/hly3) (at a concentration of 4.8×10^6 CFU for 48 h). The cells were washed and 180 μ L of medium and 20 μ L of MTT (5 mg MTT/mL solution) were added to each well. The plates were incubated for 3 h, the cells were washed and 200 μ L of DMSO was added to each well, and the formazan was solubilized on a shaker for 15 min at 150 rpm[4]. The absorbance of each well was read on a microplate reader at a test wavelength of 492 nm. The inhibition (%) of cell proliferation was determined as follows: inhibitory growth (1- Abs₄₉₂ treated cells/Abs₄₉₂ control cells) x 100% [24]. All observations were validated by three independent experiments in triplicates for each experiment.

2.7 Flow cytometry

The Guava Nexin assay (Guava Technologies) was conducted following the manufacturer's instructions. Briefly, 2.0×10^4 to 1.0×10^6 of the treated 5637 cells (100 μ L) was added to the Guava Nexin reagent. Cells were incubated in the dark at room temperature for 20 min and samples were acquired on flow cytometry Guava EasyCyte System. In this assay, an annexin V-negative and 7-AAD-positive result indicated nuclear debris, an annexin V-positive and 7-AAD-positive result indicated late apoptotic cells, while an annexin V-negative and 7-AAD-negative result indicated live healthy cells and annexin V-positive and 7-AAD-negative result indicated the presence of early apoptotic cells.

2.8 Data analysis

Data sets were analyzed using one-way ANOVA. Significance was considered at $P < 0.05$ in all analyses. Data were expressed as mean $\pm$ SEM.

3. RESULTS

3.1 Construction of rBCG*Δ*leuD

The fragment amplified by PCR, containing the entire coding sequence of the *hly*, plus the Ag85B signal peptide was cloned into pUP410 cloning vector. The fragment of 2.371 bp was successfully cloned. The insert was fully characterized by restriction enzyme digestion.

3.2 RT-PCR detection the mRNA

The presence of Hly mRNA was confirmed by RT-PCR, and the amplicon size (200bp) was confirmed by agarose gel electrophoresis at a concentration of 0.8% (Fig 2).

3.3 Determination of cytotoxicity

Human bladder carcinoma cells were incubated with the BCG strains (BCG Pasteur, BCG $\Delta leuD$, BCG $\Delta leuD/hly1$, BCG $\Delta leuD/hly2$ and BCG $\Delta leuD/hly3$) for 48 h. As demonstrated in Fig. 3, the recombinants BCG/ $\Delta leuD$ strains (BCG $\Delta leuD/hly1$, BCG $\Delta leuD/hly2$ and BCG $\Delta leuD/hly3$) did not demonstrate a significant *in vitro* cytotoxic activity after 48 h of treatment. The treatment with BCG/ $\Delta leuD$ Pasteur induced cell growth of 19%, for BCG/ $\Delta leuD$ treatment the growth inhibition was 16% and for BCG $\Delta leuD/hly1$, BCG $\Delta leuD/hly2$ and BCG $\Delta leuD/hly3$ treatments it was 18%, 19% and 14% respectively (Fig 3). This percentage of growth inhibition is not different from the BCG Pasteur control.

3.4 Flow Cytometry

The annexin-PE staining assay was performed to characterize treatments with the recombinant BCG inducing apoptosis in 5637 cell line after 48h of exposure. Annexin V binds to those cells that express phosphatidylserine on the outer layer of the cell membrane, a characteristic feature of cells entering apoptosis. The results showed that the BCG $\Delta leuD/hly$ treatments were little effective in causing cell death through apoptosis. BCG $\Delta leuD$ strain showed an early apoptosis ratio of 3.18%, while BCG $\Delta leuD/hly1$, BCG $\Delta leuD/hly2$ and BCG $\Delta leuD/hly3$ showed 3.64%, 3.70% and 3.13% of early apoptosis respectively (Fig4). For late apoptosis, the percentage ratio of

BCG Δ leuD treatment was 1.93%, for BCG Δ leuD/hly1 it was 2.16%, for BCG Δ leuD/hly2 it was 2.26% and for BCG Δ leuD/hly3 it was 2.12%. There are no statistical differences between control and BCG strains treatment.

4. DISCUSSION

The *Mycobacterium bovis* strain Bacillus Calmette-Guerin (BCG) is used as a vaccine against tuberculosis [3], is one of the most widely administered vaccines worldwide [10]. Moreover, *M. bovis* BCG-based therapy is also the treatment of choice for superficial bladder cancer [18]. Another way to improve BCG is by introducing genetic modifications for superior targeting of essential immune pathways [11]. Studies demonstrated that rBCG has a great potential as a vaccine vector, and a recombinant vaccine able to protect against tuberculosis and several other diseases is conceivable [3]. Recently, a work demonstrated an overexpression with Ag85B in a BCG Pasteur strain, using an expression system based on the use of a auxotrophic strain for the leucine amino acid, and complemented with *leuD* inserted into the plasmid vector (pUP410). This recombinant BCG strain was evaluated as a vaccine against bovine tuberculosis, and showed protection against a *M. bovis* challenge at higher level than its parental strain [14,12]. This recombinant strain demonstrated a significant in vitro cytotoxic activity after 48 h of treatment, inhibiting more than 50% of tumour cells. The inhibition of cell proliferation following BCG Δ leuD/Ag85B treatment was 77.8%, while for the other strains it was 28% (BCG Δ leuD) and 38.1% (BCG Pasteur) [4].

Intensive research has been carried out to find less toxic and more potent therapeutic agents for the immunotherapy of bladder tumors [22]. In a previous study, *Listeria monocytogenes*-based vaccines limited autochthonous tumor growth and broke immunologic tolerance in human papillomavirus [17,21]. A Molecular strategy for

some types of cancer treatment is the use of a important protein of the bacterium *L. monocytogenes*, called listeriolysin, which acts forming pores on the membrane of the early phagosome and allows leakage of the BCG immunogen from the phagosome into the cytoplasm of infected host cells[8,15]. Several groups have shown recombinant *L. monocytogenes* to be an effective agent for immunotherapy against infection and cancer [15].

In the present study, we successfully performed the cloning of the *hly* gene into BCG Pasteur $\Delta leuD$ using pUP410/*hly* as cloning vector. The transcription of the synthetic gene was confirmed by RT-PCR. BCG $\Delta leuD/hly$ liquid culture (OD₆₀₀) was used in the bacterial preparation and it was incubated with bladder cancer cells (5637 line) to conduct a preliminary test of cytotoxicity level. The initial results showed that there was no statistical difference between BCG $\Delta leuD/hly$ and conventional treatment using BCG Pasteur for this cell line. In a previous work, rBCG $\Delta leuD$ overexpressing Ag85B demonstrated enhanced cytotoxicity on superficial bladder cancer cells in vitro and this effect may be due to apoptosis and cell-cycle arrest [4]. These data corroborate that the development of immunotherapy based on recombinant BCG (rBCG) overexpressing antigens or expressing foreign antigens is a promising approach to improve the performance of BCG anti-tumour therapy [2].

The listeriolysin expressed in rBCG $\Delta leuD/hly$ can allow access to both phagosomal and cytosolic compartments and BCG antigens may be presented in the context of both MHC I and II molecules, resulting in strong cellular immune responses [8]. Previous studies have indicated that when fused to a truncated form of listeriolysin, antigens expressed and secreted are significantly more immunogenic than when they are secreted as stand-alone proteins [9,16]. Moreover, a recombinant *L. monocytogenes*, expressing and secreting the human PSA protein fused to Hly, was able to generate an

effective and durable anti-tumor response in mice [15]. Based on this, we constructed a new recombinant BCG strain, comprised of an rBCG Δ leuD which expresses and secretes the Hly. Our preliminary evaluation demonstrated that the rBCG strain had no enhanced cytotoxic activity on bladder cancer cells hly, however, this strain may be evaluated in other assay using other bladder cell lines or different kinds of cancer, and the result might be different.

Tumor suppression frequently involves the modulation of signal transduction pathways, leading to alterations in gene expression, arrest in cell cycle progression or apoptosis [23]. Apoptosis is considered a highly regulated process that allows a cell to self-degrade in order to eliminate an unwanted or dysfunctional cell [20]. Anticancer treatments kill tumor cells primarily by the induction of apoptosis through activation of the intrinsic pathway [20]. In this work, we performed flow cytometry assay to verify apoptosis level in bladder cancer cell. The results showed that the level of apoptosis induced by BCG Δ leuD/hly was similar to BCG Pasteur employed on the conventional therapy in bladder cancer cells. There were no significant differences in comparison to the control group.

In conclusion, the preliminary data here presented indicate that a recombinant BCG Δ leuD/hly, can generate an anti-tumor response similar to the BCG Pasteur in the treatment of bladder cancer cells. Our findings indicate that it maintains the same cytotoxicity levels in vitro. Further investigation of the rBCG strain may be evaluated as a new immunotherapeutic agent against other kinds of cancer.

References

1. Ali D, Ray RS, Hans RK (2010) UVA-induced cytotoxicity and DNA damaging potential of benz (e) acephenanthrylene. *Toxicol Lett* 199:193-200
2. Amirkhah R, Khanahmad H, Abolhassani M et al (2009) Improvement of bladder cancer immunotherapy by creating a recombinant Bacille Calmette-Gu'erin which secretes p53 protein. *Med Hypotheses* 72:754-
3. Bastos RG, Borsuk S, Seixas FK et al (2009) Recombinant Mycobacterium bovis BCG. *Vaccine* 27:6495-6503
4. Begnini KR, Rizzi C, Campos VF et al (2013) Auxotrophic recombinant Mycobacterium bovis BCG overexpressing Ag85B enhances cytotoxicity on superficial bladder cancer cells in vitro. *Appl Microbiol Biotechnol* 97:1543-1552
5. Borsuk S, Mendum TA, Fagundes MQ et al (2007) Auxotrophic complementation as a selectable marker for stable expression of foreign antigens in Mycobacterium bovis BCG. *Tuberculosis (Edinb)* 87:474-480
6. Dennehy M, Williamson AL (2005) Factors influencing the immune response to foreign antigen expressed in recombinant BCG vaccines. *Vaccine* 23:1209-1224
7. Ding GQ, Yu YL, Shen ZJ et al (2012) Antitumor effects of human interferon-alpha 2b secreted by recombinant bacillus Calmette-Guerin vaccine on bladder cancer cells. *J Zhejiang Univ Sci B* 13:335-341
8. Grode L, Seiler P, Baumann S et al (2005) Increased vaccine efficacy against tuberculosis of recombinant Mycobacterium bovis bacille Calmette-Guerin mutants that secrete listeriolysin. *J Clin Invest* 115:2472-2479
9. Gunn GR, Zubair A, Peters C et al (2001) Two Listeria monocytogenes vaccine vectors that express different molecular forms of human papilloma virus-16 (HPV-16) E7 induce qualitatively different T cell immunity that correlates with their ability to induce regression of established tumors immortalized by HPV-16. *J Immunol* 167:6471-6479
10. Hesselning AC, Marais BJ, Gie RP et al (2007) The risk of disseminated Bacille Calmette-Guerin (BCG) disease in HIV-infected children. *Vaccine* 25:14-18
11. Reece ST, Kaufmann SH (2012) Floating between the poles of pathology and protection: can we pin down the granuloma in tuberculosis? *Curr Opin Microbiol* 15:63-70
12. Rizzi C, Bianco MV, Blanco FC et al (2012) Vaccination with a BCG strain overexpressing Ag85B protects cattle against Mycobacterium bovis challenge. *PLoS One* 7:e51396-

13. Schaible UE, Winau F, Sieling PA et al (2003) Apoptosis facilitates antigen presentation to T lymphocytes through MHC-I and CD1 in tuberculosis. *Nat Med* 9:1039-1046
14. Seixas FK, Borsuk S, Fagundes MQ et al (2010) Stable expression of *Leptospira interrogans* antigens in auxotrophic *Mycobacterium bovis* BCG. *Biol Res* 43:13-18
15. Shahabi V, Reyes-Reyes M, Wallecha A et al (2008) Development of a *Listeria monocytogenes* based vaccine against prostate cancer. *Cancer Immunol Immunother* 57:1301-1313
16. Singh R, Wallecha A (2011) Cancer immunotherapy using recombinant *Listeria monocytogenes*: transition from bench to clinic. *Hum Vaccin* 7:497-505
17. Souders NC, Sewell DA, Pan ZK et al (2007) *Listeria*-based vaccines can overcome tolerance by expanding low avidity CD8+ T cells capable of eradicating a solid tumor in a transgenic mouse model of cancer. *Cancer Immun* 7:2-
18. Sylvester RJ, van der Meijden AP, Lamm DL (2002) Intravesical bacillus Calmette-Guerin reduces the risk of progression in patients with superficial bladder cancer: a meta-analysis of the published results of randomized clinical trials. *J Urol* 168:1964-1970
19. Triccas JA (2010) Recombinant BCG as a vaccine vehicle to protect against tuberculosis. *Bioeng Bugs* 1:110-115
20. Vangestel C, Van de Wiele C, Mees G et al (2009) Forcing cancer cells to commit suicide. *Cancer Biother Radiopharm* 24:395-407
21. Wallecha A, Maciag PC, Rivera S et al (2009) Construction and characterization of an attenuated *Listeria monocytogenes* strain for clinical use in cancer immunotherapy. *Clin Vaccine Immunol* 16:96-103
22. Yuksel ZS, Buber E, Kocagoz T et al (2011) *Mycobacterial* strains that stimulate the immune system most efficiently as candidates for the treatment of bladder cancer. *J Mol Microbiol Biotechnol* 20:24-28
23. Yun JM, Kweon MH, Kwon H et al (2006) Induction of apoptosis and cell cycle arrest by a chalcone panduratin A isolated from *Kaempferia pandurata* in androgen-independent human prostate cancer cells PC3 and DU145. *Carcinogenesis* 27:1454-1464
24. Zheng D, Wang Y, Zhang D et al (2011) In vitro antitumor activity of silybin nanosuspension in PC-3 cells. *Cancer Lett* 307:158-164

Figures

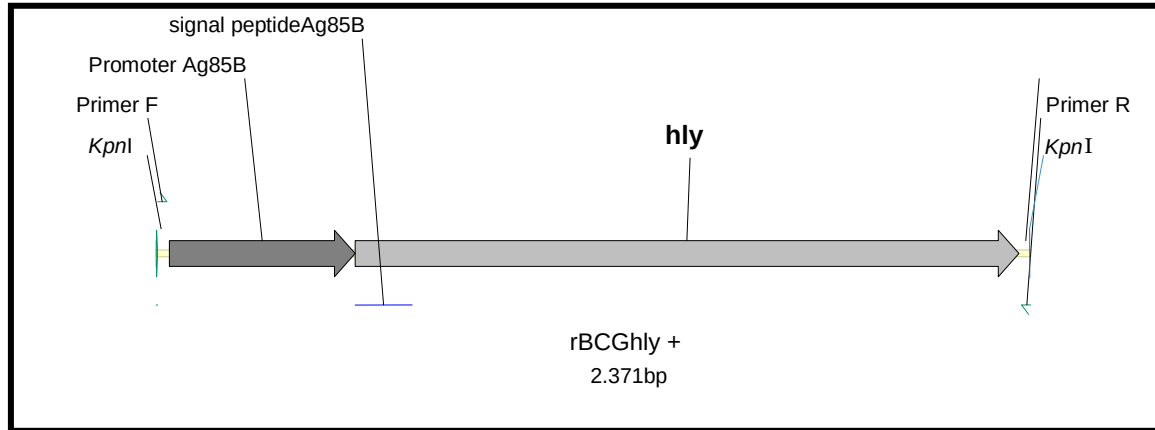


Figure 1. Schematic representation of the synthetic gene cloned in vector pUP410.

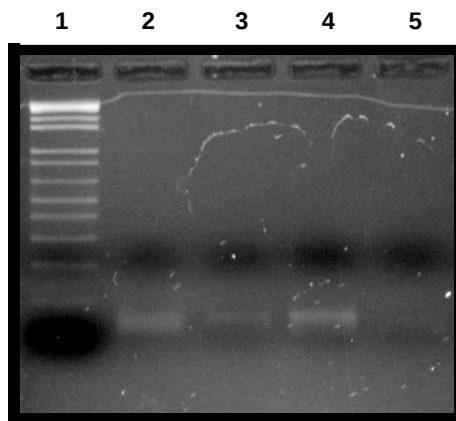


Figure 2.(A) Electrophoresis in a 0.8% agarose gel. Lane 1, 1 kb Plus DNA Ladder (Invitrogen®); Lane 2, rBCG/hly1; Lane 3, rBCG/hly2; Lane 4, rBCG/hly3; and Lane 5, negative control;

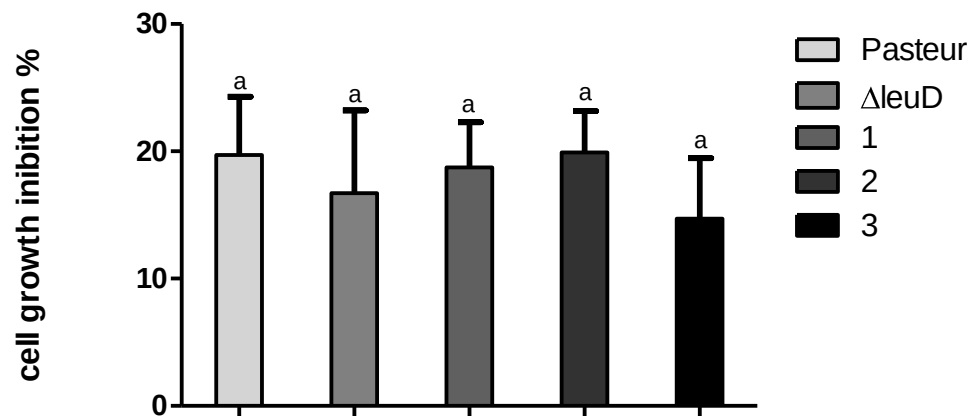


Figure 3. Human bladder carcinoma cells (5637) growth inhibited after 48h of treatment with BCG $\Delta leuD/hly$. Cell proliferation was investigated by MTT assay. Data are expressed as means $\pm$ SEM from three independent experiments.

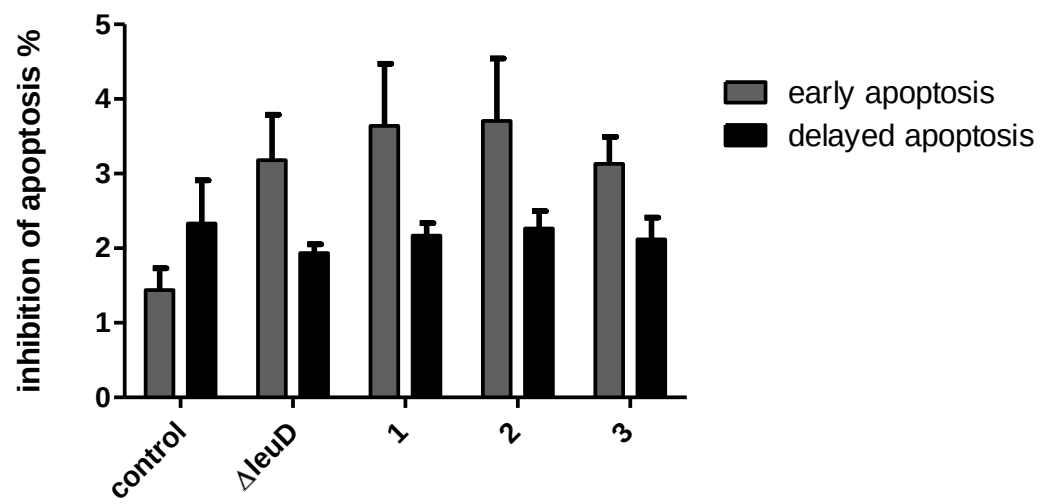


Figure 4. Inhibition rate of early and delayed apoptosis, in treated BCG/ ΔeuD /hly; data are expressed as means $\pm$ SEM from three independent experiments.

4. CONCLUSÕES

- A cepa auxotrófica BCG Δ *euD*/*hly* apresenta um nível de citotoxicidade similar à cepa BCG Pasteur utilizada no tratamento convencional;
- A expressão do gene que codifica para a listeriolisina não confere um aumento na atividade citotóxica do BCG em linhagem celular de câncer de bexiga, conforme era esperado.

5.REFERÊNCIAS

ALDRICH, J. F.; LOWE, D. B.; SHEARER, M. H.; WINN, R. E.; JUMPER, C. A.; KENNEDY, R. C. Vaccines and immunotherapeutics for the treatment of malignant disease. **Clin.Dev.Immunol.**, v.2010, p.697158, 2010.

AMIRKHAH, R.; KHANAHMAD, H.; ABOLHASSANI, M.; POOYA, M.; MOVASSAGH, H.; SHOKRGOZAR, M. A. Improvement of bladder cancer immunotherapy by creating a recombinant Bacille Calmette-Gu'erin which secretes p53 protein. **Med.Hypotheses**, v.72, n.6, p.754, 2009.

ALI D, RAY RS, HANS RK (2010) UVA-induced cytotoxicity and DNA damaging potential of benz (e) acephenanthrylene. **Toxicol Lett** 199:193-200.

ANDRADE, P. M.; CHADE, D. C.; BORRA, R. C.; NASCIMENTO, I. P.; VILLANOVA, F. E.; LEITE, L. C.; ANDRADE, E.; SROUGI, M. The therapeutic potential of recombinant BCG expressing the antigen S1PT in the intravesical treatment of bladder cancer. **Urol.Oncol.**, v.28, n.5, p.520-525, 2010.

BANGE, F. C.; BROWN, A. M.; JACOBS, W. R., JR. Leucine auxotrophy restricts growth of Mycobacterium bovis BCG in macrophages. **Infect.Immun.**, v.64, n.5, p.1794-1799, 1996.

BASTOS, R. G.; BORSUK, S.; SEIXAS, F. K.; DELLAGOSTIN, O. A. Recombinant *Mycobacterium bovis* BCG. **Vaccine**, v.27, n.47, p.6495-6503, 2009.

BEGINI, K. R.; RIZZI, C.; CAMPOS, V. F.; BORSUK, S.; SCHULTZE, E.; YURGEL, V. C.; NEDEL, F.; DELLAGOSTIN, O. A.; COLLARES, T.; SEIXAS, F. K. Auxotrophic recombinant *Mycobacterium bovis* BCG overexpressing Ag85B enhances cytotoxicity on superficial bladder cancer cells in vitro. **Appl.Microbiol.Biotechnol.**, v.97, n.4, p.1543-1552, 2013.

BORSUK, S.; MENDUM, T. A.; FAGUNDES, M. Q.; MICHELON, M.; CUNHA, C. W.; MCFADDEN, J.; DELLAGOSTIN, O. A. Auxotrophic complementation as a selectable marker for stable expression of foreign antigens in *Mycobacterium bovis* BCG. **Tuberculosis.(Edinb.)**, v.87, n.6, p.474-480, 2007.

CHUNG, M. A.; LUO, Y.; O'DONNELL, M.; RODRIGUEZ, C.; HEBER, W.; SHARMA, S.; CHANG, H. R. Development and preclinical evaluation of a *Bacillus Calmette-Guerin*-MUC1-based novel breast cancer vaccine. **Cancer Res.**, v.63, n.6, p.1280-1287, 2003.

DRANOFF, G. Cytokines in cancer pathogenesis and cancer therapy. **Nat.Rev.Cancer**, v.4, n.1, p.11-22, 2004.

DENNEHY M, WILLIAMSON AL (2005) Factors influencing the immune response to foreign antigen expressed in recombinant BCG vaccines. *Vaccine* 23:1209-1224.

DING GQ, YU YL, SHEN ZJ et al (2012) Antitumor effects of human interferon-alpha 2b secreted by recombinant bacillus Calmette-Guerin vaccine on bladder cancer cells. *J Zhejiang Univ Sci B* 13:335-341.

FORRELLAD, M. A.; KLEPP, L. I.; GIOFFRE, A.; GARCIA, J.; MORBIDONI, H. R.; DE LA PAZ, S. M.; CATALDI, A. A.; BIGI, F. Virulence factors of the *Mycobacterium tuberculosis* complex. **Virulence.**, v.4, n.1, p.3-66, 2013.

GRODE L, SEILER P, BAUMANN S et al (2005) Increased vaccine efficacy against tuberculosis of recombinant *Mycobacterium bovis* bacille Calmette-Guerin mutants that secrete listeriolysin. *J Clin Invest* 115:2472-2479.

GUNN GR, ZUBAIR A, PETERS C et al (2001) Two *Listeria monocytogenes* vaccine vectors that express different molecular forms of human papilloma virus-16 (HPV-16) E7 induce qualitatively different T cell immunity that correlates with their ability to induce regression of established tumors immortalized by HPV-16. *J Immunol* 167:6471-6479.

HESSELING AC, MARAIS BJ, GIE RP et al (2007) The risk of disseminated Bacille Calmette-Guerin (BCG) disease in HIV-infected children. *Vaccine* 25:14-18.

PARISH, T.; STOKER, N. G. Electroporation of mycobacteria. **Methods Mol.Biol.**, v.101, p.129-144, 1998.

REECE ST, KAUFMANN SH (2012) Floating between the poles of pathology and protection: can we pin down the granuloma in tuberculosis? *Curr Opin Microbiol* 15:63-70.

RIZZI, C.; BIANCO, M. V.; BLANCO, F. C.; SORIA, M.; GRAVISACO, M. J.; MONTENEGRO, V.; VAGNONI, L.; BUDDLE, B.; GARBACCIO, S.; DELGADO, F.; LEAL, K. S.; CATALDI, A. A.; DELLAGOSTIN, O. A.; BIGI, F. Vaccination with a BCG strain overexpressing Ag85B protects cattle against *Mycobacterium bovis* challenge. **PLoS.One.**, v.7, n.12, p.e51396, 2012.

SCHAIBLE UE, WINAU F, SIELING PA et al (2003) Apoptosis facilitates antigen presentation to T lymphocytes through MHC-I and CD1 in tuberculosis. *Nat Med* 9:1039-1046.

SHAHABI, V.; REYES-REYES, M.; WALLECHA, A.; RIVERA, S.; PATERSON, Y.; MACIAG, P. Development of a *Listeria monocytogenes* based vaccine against prostate cancer. **Cancer Immunol.Immunother.**, v.57, n.9, p.1301-1313, 2008.

SEIXAS FK, BORSUK S, FAGUNDES MQ et al (2010) Stable expression of *Leptospira interrogans* antigens in auxotrophic *Mycobacterium bovis* BCG. *Biol Res* 43:13-18.

SINGH R, WALLECHA A (2011) Cancer immunotherapy using recombinant *Listeria monocytogenes*: transition from bench to clinic. *Hum Vaccin* 7:497-505.

SOUDERS NC, SEWELL DA, PAN ZK et al (2007) *Listeria*-based vaccines can overcome tolerance by expanding low avidity CD8⁺ T cells capable of eradicating a solid tumor in a transgenic mouse model of cancer. *Cancer Immun* 7:2.

SYLVESTER RJ, VAN DER MEIJDEN AP, LAMM DL (2002) Intravesical bacillus Calmette-Guerin reduces the risk of progression in patients with superficial bladder cancer: a meta-analysis of the published results of randomized clinical trials. *J Urol* 168:1964-1970.

TRICCAS JA (2010) Recombinant BCG as a vaccine vehicle to protect against tuberculosis. *Bioeng Bugs* 1:110-115.

VANGESTEL C, VAN DE WIELE C, MEES G et al (2009) Forcing cancer cells to commit suicide. *Cancer Biother Radiopharm* 24:395-407.

VARALDO, P. B.; LEITE, L. C.; DIAS, W. O.; MIYAJI, E. N.; TORRES, F. I.; GEBARA, V. C.; ARMOA, G. R.; CAMPOS, A. S.; MATOS, D. C.; WINTER, N.; GICQUEL, B.; VILAR, M. M.; MCFADDEN, J.; ALMEIDA, M. S.; TENDLER, M.; MCINTOSH, D. Recombinant *Mycobacterium bovis* BCG expressing the Sm14 antigen of *Schistosoma mansoni* protects mice from cercarial challenge. **Infect.Immun.**, v.72, n.6, p.3336-3343, 2004.

VON REYN, C. F.; VUOLA, J. M. New vaccines for the prevention of tuberculosis. **Clin.Infect.Dis.**, v.35, n.4, p.465-474, 2002.

WANG, C.; FU, R.; CHEN, Z.; TAN, K.; CHEN, L.; TENG, X.; LU, J.; SHI, C.; FAN, X. Immunogenicity and protective efficacy of a novel recombinant BCG strain overexpressing antigens Ag85A and Ag85B. **Clin.Dev.Immunol.**, v.2012, p.563838, 2012.

WALLECHA A, MACIAG PC, RIVERA S et al (2009) Construction and characterization of an attenuated *Listeria monocytogenes* strain for clinical use in cancer immunotherapy. *Clin Vaccine Immunol* 16:96-103.

YUKSEL ZS, BUBER E, KOCAGOZ T et al (2011) Mycobacterial strains that stimulate the immune system most efficiently as candidates for the treatment of bladder cancer. *J Mol Microbiol Biotechnol* 20:24-28.

YUN JM, KWEON MH, KWON H et al (2006) Induction of apoptosis and cell cycle arrest by a chalcone panduratin A isolated from *Kaempferia pandurata* in androgen-independent human prostate cancer cells PC3 and DU145. *Carcinogenesis* 27:1454-1464.

ZHENG D, WANG Y, ZHANG D et al (2011) In vitro antitumor activity of silybin nanosuspension in PC-3 cells. *Cancer Lett* 307:158-164.